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# Position of Side-chain Branching and Handedness of Turns and Helices of Homopeptides from Chiral $C^{\alpha}$ -Methylated Amino Acids. Crystal-state Structural Analysis of ( $\alpha$ Me)Leu Trimer and Tetramer

André Aubry,<sup>a</sup> Daniel Bayeul,<sup>a</sup> Gilles Précigoux,<sup>b</sup> Monica Pantano,<sup>c</sup> Fernando Formaggio,<sup>c</sup> Marco Crisma,<sup>c</sup> Claudio Toniolo,<sup>c</sup> Wilhelmus H. J. Boesten,<sup>d</sup> Hans E. Schoemaker<sup>d</sup> and Johan Kamphuis<sup>d</sup>

<sup>a</sup> Laboratory of Mineralogy and Crystallography, University of Nancy I, 54506 Vandoeuvre, France

<sup>b</sup> Laboratory of Crystallography and Crystalline Physics, University of Bordeaux I, 33405 Talence, France

<sup>c</sup> Biopolymer Research Čentre, CNR, Department of Organic Chemistry, University of Padova,

35131 Padova, Italy

<sup>d</sup> DSM Research, Bio-organic Chemistry Section, 6160 MD Geleen, The Netherlands

Terminally blocked homotri- and homotetra-peptides from  $(\alpha Me)Leu$ , a chiral  $C_{\alpha}$ -methylated,  $\gamma$ -branched  $\alpha$ -amino acid, have been prepared by solution methods and fully characterized. The molecular and crystal structures of  $\rho$ BrBz-[D- $(\alpha Me)Leu]_3$ -OH monohydrate and  $\rho$ BrBz-[D- $(\alpha Me)-Leu]_4$ -OBu<sup>t</sup> (where  $\rho$ BrBz indicates  $\rho$ -bromobenzoyl) were determined by X-ray diffraction. The tripeptide carboxylic acid adopts a type-III  $\beta$ -turn conformation followed by an uncommon oxy-analogue of a type-III  $\beta$ -turn, the latter being stabilized by a 1 $\leftarrow$ 4 C=O···H–O intramolecular H-bond. The three independent molecules in the asymmetric unit of the tetrapeptide ester are folded in a regular right-handed  $3_{10}$ -helix. All ( $\alpha$ Me)Leu residues exhibit  $\varphi$ ,  $\psi$  torsion angles in the helical region of the conformational map. These results indicate that: (*i*) the ( $\alpha$ Me)Leu residue is an effective  $\beta$ -turn and helix promoter and (*ii*) the relationship between ( $\alpha$ Me)Leu chirality and turn and helix handedness is the same as that shown by the  $\gamma$ -branched ( $\alpha$ Me)Phe residue, but it is opposite to that characteristic of isovaline (Iva), with a linear side chain, the  $\beta$ -branched ( $\alpha$ Me)Val residue and protein amino acids (including Leu).

The stereochemistry of peptides containing  $C_{\alpha}$ -methylated  $\alpha$ -amino acids is rather unique as they possess significant constraints on their conformational freedom.<sup>1</sup> This property is of great relevance to the exploitation of these compounds as: (*i*) precise molecular rulers or as scaffolding units in the *de novo* design of protein and enzyme mimetics and in the investigation of molecular recognition processes,<sup>2-4</sup> and (*ii*) conformation-ally restricted, enzyme-resistant agonists and antagonists of bioactive peptides.<sup>5-7</sup>

Conformational energy computations, pioneered by Marshall in the early '70s,<sup>1</sup> showed that the presence of a *gem*-dimethyl group on the  $\alpha$ -carbon of glycine imposes a marked restriction on the available conformational space of the resulting amino acid (Aib,  $\alpha$ -aminoisobutyric acid or  $C_{\alpha}$ -methylalanine) (Scheme 1). X-Ray diffraction analyses unequivocally established that only the type-III (III')  $\beta$ -turn<sup>8-10</sup> and the regular right- (and left-) handed  $3_{10}$ -helix<sup>11</sup> are adopted by the achiral Aib homo-oligomers (to the decamer) in the crystal state.<sup>3,12-15</sup>

In our continuing investigation of the crystal-state preferred conformation of homopeptides from  $C_{\alpha}$ -methylated  $\alpha$ -amino acids, we have recently been able to solve the X-ray structures of a variety of homochiral homopeptides from the Iva (isovaline or  $C_{\alpha}$ -methyl- $\alpha$ -aminobutyric acid), ( $\alpha$ Me)Val ( $C_{\alpha}$ -methylvaline) and ( $\alpha$ Me)Phe ( $C_{\alpha}$ -methylphenylalanine) residues.<sup>16</sup> The types of secondary structure observed are the same as those described above for Aib ( $\beta$ -turns and  $3_{10}$ -helices). However, the relationship between  $\alpha$ -carbon chirality and turn and helix screw sense strongly depends upon presence and position of side-chain branching. Specifically, Iva, with a linear side chain, and the  $\beta$ -branched, aliphatic ( $\alpha$ Me)Val exhibit a normal behaviour (the same as that shown by protein amino acids), while for the  $\gamma$ -branched, aromatic ( $\alpha$ Me)Phe the relationship is inverse.

With the aim of defining more accurately the role played by



Scheme 1  $\alpha$ -Amino acids methylated at the  $\alpha$ -carbon discussed in this work

position of side-chain branching and aromaticity, we report here a crystal-state structural study by X-ray diffraction of the trimer and tetramer from ( $\alpha$ Me)Leu, a  $C_{\alpha}$ -methylated amino acid with a  $\gamma$ -branched, aliphatic side chain.

#### Experimental

*Materials.*—The synthesis and characterization of the two ( $\alpha$ Me)Leu homochiral homo-oligomers, the X-ray diffraction structures of which are discussed in this work, are reported below, while those of other members of the series (to the tetramer level) will be described elsewhere.<sup>17</sup>



Fig. 1 X-Ray diffraction structure of  $pBrBz-[D-(\alpha Me)Leu]_3$ -OH monohydrate with atom numbering. The two intramolecular H-bonds are indicated as dashed lines.

pBrBz-[D-(αMe)Leu]<sub>3</sub>-OH (pBrBz, p-bromobenzoyl).—This compound was prepared by stirring pBrBz-[D-(αMe)Leu]<sub>3</sub>-OBu<sup>t</sup> in a 1:1 trifluoroacetic acid–dichloromethane mixture at room temperature for 2 h. The solvent was removed *in vacuo* and the residue evaporated several times from diethyl ether. Yield 80%. M.p. 219–220 °C (from ethyl acetate–light petroleum);  $[\alpha]_{D}^{20}$  – 5.8 (*c* 0.5 in methanol); TLC (silica gel plates 60F-254, Merck)  $R_{F1}$  (chloroform–ethanol 9:1) 0.40,  $R_{F2}$ (butan-1-ol–acetic acid–water 3:1:1) 0.95,  $R_{F3}$  (toluene–ethanol 7:1) 0.25;  $v_{max}(1 \text{ mmol dm}^{-3} \text{ CDCl}_3)/\text{cm}^{-1}$  3505, 3450, 3407, 3363, 1741, 1671 and 1649;  $\delta_{H}(220 \text{ MHz}; 10 \text{ mmol dm}^{-3}$ CDCl<sub>3</sub>; Me<sub>4</sub>Si) 7.68 and 7.57 (2 m, 4 H, *p*BrBz CH), 7.53 (s, 1 H, NH), 7.25 (s, 1 H, NH), 7.10 (s, 1 H, NH), 2.50–2.23 (m, 3 H, 3 γ-CH), 1.90–1.50 (m, 6 H, 3β-CH<sub>2</sub>), 1.70 (s, 3 H, β-CH<sub>3</sub>), 1.64 (s, 3 H, β-CH<sub>3</sub>), 1.60 (s, 3 H, β-CH<sub>3</sub>) and 0.94–0.82 (m, 18 H, 6 δ-CH<sub>3</sub>) (Found: C, 56.9; H, 7.6; N, 7.1. Calc. for C<sub>28</sub>H<sub>44</sub>BrN<sub>3</sub>O<sub>5</sub>: C, 57.7; H, 7.6; N, 7.2%).

pBrBz-[D-(aMe)Leu]<sub>4</sub>-OBu<sup>t</sup>.—This compound was synthesized from the oxazol-5(4H)-one from  $pBrBz-[D-(\alpha Me)Leu]_3$ -OH and H-D-( $\alpha$ Me)Leu-OBu<sup>t</sup> in acetonitrile under reflux for 43 h. The solvent was removed under reduced pressure, the residue was dissolved in ethyl acetate and the organic solution was washed with 10% KHSO<sub>4</sub>, water, 5% NaHCO<sub>3</sub>, water, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and evaporated to dryness. The product was purified on a silica gel column eluted with a stepwise gradient of ethyl acetate in toluene. Yield 36%. M.p. 220–222 °C (from diethyl ether–light petroleum);  $[\alpha]_{\rm D}^{20}$  31.4 (c 0.5 in methanol); TLC  $R_{F1}$  0.95,  $R_{F2}$  0.95,  $R_{F3}$  0.70;  $\nu_{max}(1 \text{ mmol } dm^{-3} \text{ CDCl}_3)/cm^{-1}$  3389, 3364, 1716, 1676 and 1648;  $\delta_{\rm H}(10 \text{ mmol dm}^{-3} \text{ CDCl}_3)$  7.97 (s, 1 H, NH), 7.71 and 7.55 (2 m, 4 H, pBrBz CH), 7.67 (s, 1 H, NH), 7.61 (s, 1 H, NH), 7.03 (s, 1 H, NH), 2.80, 2.63 and 2.41 (3 m, 4 H, 4 γ-CH), 1.75-1.45  $(m, 8 H, 4 \beta$ -CH<sub>2</sub>), 1.74 (s, 3 H,  $\beta$ -CH<sub>3</sub>), 1.64 (s, 3 H,  $\beta$ -CH<sub>3</sub>), 1.62 (s, 3 H, β-CH<sub>3</sub>), 1.55 (s, 3 H, β-CH<sub>3</sub>), 1.51 (s, 9 H, 3 OBu<sup>t</sup>CH<sub>3</sub>) and 0.89–0.81 (m, 24 H, 8 δ-CH<sub>3</sub>) (Found: C, 61.1; H, 8.7; N, 7.2. Calc. for C<sub>39</sub>H<sub>65</sub>BrN<sub>4</sub>O<sub>6</sub>: C, 61.2; H, 8.6; N, 7.3%).

Crystallographic Data for pBrBz-[D-( $\alpha$ Me)Leu]<sub>3</sub>-OH monohydrate.—C<sub>28</sub>H<sub>46</sub>BrN<sub>3</sub>O<sub>6</sub>, M = 600.6. Orthorhombic, a = 9.805(2), b = 17.453(3), c = 19.535(3) Å, V = 3343 Å<sup>3</sup>, space group P2<sub>1</sub>2<sub>1</sub>2<sub>1</sub>, Z = 4,  $D_c = 1.19$  g cm<sup>-3</sup>, F(000) = 1272,  $\mu = 18.2$  cm<sup>-1</sup> (Cu-K $\alpha$ ), final R value 0.086. Crystallographic Data for pBrBz-[D-( $\alpha$ Me)Leu]<sub>4</sub>-OBu<sup>t</sup>.— C<sub>39</sub>H<sub>65</sub>BrN<sub>4</sub>O<sub>6</sub>, M = 765.9. Trigonal, a = b = 19.583(3), c = 30.459(4) Å,  $V = 10\,116$  Å<sup>3</sup>, space group P3<sub>2</sub>, Z = 9,  $D_c = 1.03$  g cm<sup>-3</sup>, F(000) = 3690,  $\mu = 14.66$  cm<sup>-1</sup> (Cu-K $\alpha$ ), final R value 0.111.

X-Ray Crystal Structure Determination of pBrBz-[D-( $\alpha$ Me)-Leu]<sub>3</sub>-OH monohydrate and pBrBz-[D-(aMe)Leu]<sub>4</sub>-OBu<sup>t</sup>.-Colourless crystals of the trimer and tetramer were grown by slow evaporation of a methanol-water solvent mixture and a methanol solution, respectively. Enraf-Nonius CAD4 diffractometer,  $\omega/2\theta$  scan mode up to  $\theta = 60^{\circ}$  for the trimer and  $\theta = 70^{\circ}$  for the tetramer; graphite-monochromated Cu-K<sub> $\alpha$ </sub> radiation ( $\lambda = 1.5418$  Å); 2813 and 12 390 unique reflections for the trimer and tetramer, respectively; 1397 and 5854 reflections with  $I \ge 1 \sigma(I)$  considered observed for the trimer and tetramer, respectively. The two structures were solved by SHELXS 8618 and refined by blocked least squares with  $w = 0.086/[\sigma^2(F) + 0.002 F^2]$  for the trimer and w = $2.219/[\sigma^2(F) + 0.0032 F^2]$  for the tetramer. The thermal parameters were anisotropic for all non-hydrogen atoms of the trimer and most of the non-hydrogen atoms of the tetramer. The trimer co-crystallized with one water molecule and exhibited a remarkable disorder at the level of the three  $(\alpha Me)$ Leu isobutyl side chains. The terminal parts of the three independent molecules in the asymmetric unit of the tetramer  $[(\alpha Me)Leu$  side chains and *tert*-butyl ester group] undergo a significant thermal motion. Some side-chain  $C_{\alpha}$  atoms were refined as staggered atoms with an occupancy factor of 2/3 and refined isotropically. Also the tert-butyl ester carbon atoms of molecule C were refined isotropically. Nevertheless, the peptide backbones are defined precisely. The hydrogen atoms of both trimer and tetramer were fixed by calculations and not refined. All calculations were performed using the SHELX 76 program.19

Fractional atomic coordinates, tables of hydrogen atoms coordinates, thermal parameters, bond lengths, bond angles, and torsion angles for the trimer and tetramer are available from the Cambridge Crystallographic Data Centre.\*

## **Results and Discussion**

We determined by X-ray diffraction the molecular and crystal structures of the following two ( $\alpha$ Me)Leu homopeptides: *p*BrBz-[D-( $\alpha$ Me)Leu]<sub>3</sub>-OH (in its monohydrate form) and *p*BrBz-[D-( $\alpha$ Me)Leu]<sub>4</sub>-OBu'. The *p*-bromobenzoyl group was incorporated at the *N*-terminus of the peptide chain to help solve the phase problem in the X-ray diffraction analyses, since it possesses a suitable heavy atom (Br). In the asymmetric unit of the tetramer three conformationally distinct, independent molecules (A, B and C) are observed. The molecular structures with the atomic numbering schemes are illustrated in Figs. 1 and 2. Relevant backbone and side-chain torsion angles<sup>20</sup> are given in Table 1. In Table 2 the intra- and inter-molecular H-bond parameters are listed.

Bond lengths and bond angles (deposited) are in general agreement with previously reported values for the geometry of the *p*-bromobenzamido moiety,<sup>21,22</sup> the *tert*-butyl ester group<sup>23</sup> and the peptide unit.<sup>24,25</sup>

All 15 D-( $\alpha$ Me)Leu residues are found in the helical region A of the conformational map.<sup>26</sup> The average absolute  $\varphi$  and  $\psi$  values are 57 and 30°, matching perfectly those expected for a  $3_{10}$ -helix.<sup>27</sup> In general, the signs of the  $\varphi$  and  $\psi$  values are negative, the only exceptions being represented by the C-terminal  $\varphi_4$  and  $\psi_4$  values of all three molecules of the tetramer.

<sup>\*</sup> For details of the CCDC deposition scheme see 'Instructions for Authors (1994),' J. Chem. Soc., Perkin Trans. 2, 1994, issue 1.



Fig. 2 X-Ray diffraction structure of the three independent molecules (A, B and C) in the asymmetric unit of  $pBrBz-[D-(\alpha Me)Leu]_4$ -OBu' with atom numbering. The two intramolecular H-bonds are indicated as dashed lines.

The  $N_{\alpha}$ -blocked tripeptide carboxylic acid forms a regular type-III  $\beta$ -turn, followed by an unusual oxy-analogue of a type-III  $\beta$ -turn.<sup>28</sup> The intramolecular N<sup>3</sup>···O<sup>0</sup> and O<sup>T</sup>···O<sup>1</sup> distances are 2.93(1) and 2.65(1) Å, respectively, within the range expected for such H-bonds.<sup>29–33</sup> The three molecules of

the  $N^{\alpha}$ -blocked tetrapeptide ester are folded in a regular, righthanded (incipient)  $3_{10}$ -helical structure, characterized by two consecutive  $1 \leftarrow 4$  C=O···H-N intramolecular H-bonds (N<sup>3</sup>···O<sup>0</sup> and N<sup>4</sup>···O<sup>1</sup>). In general, the observed N···O separations are at the upper limit for such an interaction.<sup>29-31</sup> The opposite handedness of the *C*-terminal residue of the tetrapeptide with respect to that of the preceding ones (see above) is a common observation for a  $3_{10}$ -helix forming peptide ester.<sup>34</sup>

The distribution of the  $\chi^1$  torsion angle, the most relevant for characterizing side-chain conformations, for the 15 D-( $\alpha$ Me)Leu residues is 13 g<sup>+</sup> and 2 t, in agreement with the results of statistical analyses of the Leu residue in peptides and proteins.<sup>25,35-37</sup> The disposition of residue 4 of molecules **A** and **B** of the tetramer (t) compared with that of molecule **C** (g<sup>+</sup>) represents the major conformational difference among these molecules.

All amide, peptide and ester groups ( $\omega$  torsion angles) are *trans*, as expected, with only one peptide bond ( $\omega_3$  for molecule **C** of the tetramer) deviating more than 8° from

Table 1Selected torsion angles (degrees) for  $pBrBz-[D-(\alpha Me)Leu]_3$ -<br/>OH monohydrate and  $pBrBz-[D-(\alpha Me)Leu]_4$ -OBu<sup>t</sup>

		Tetrapeptide		
	Tripeptide	Molecule A	Molecule <b>B</b>	Molecule C
θ	-20.3(19)	26.6(29)	29.6(30)	28.5(31)
ω	- 177.5(11)	-179.7(18)	-177.0(17)	-175.5(17)
Ø1	-52.3(15)	-62.3(22)	-59.2(21)	-67.0(21)
$\psi_1$	-40.1(15)	-28.9(16)	-35.5(16)	-19.5(16)
$\omega_1$	-176.6(11)	-178.4(10)	-177.5(10)	174.8(11)
$\varphi_{2}$	-46.7(16)	-61.1(15)	- 56.0(16)	- 52.9(16)
$\psi_{2}$	- 35.5(19)	-10.9(21)	-17.4(22)	-17.0(17)
$\omega_{2}$	177.9(14)	175.6(15)	178.0(14)	173.6(13)
$\tilde{\varphi_3}$	-45.5(19)	- 59.6(22)	-60.7(22)	- 56.0(23)
Ψ <sub>3</sub>	$-40.6(18)^{a}$	-26.3(24)	-25.0(24)	-23.4(26)
ω	_	-172.8(17)	-173.5(17)	- 166.5(19)
$\varphi_{\mathbf{A}}$	_	51.9(26)	53.6(26)	65.1(37)
ΨT	_	59.9(20)	48.9(20)	26.4(52)
ω	_	174.0(18)	173.9(20)	178.2(40)
$\chi_1^1$	56.3(16)	51.1(26)	56.6(25)	42.1(21)
$\chi_1^{2,1}$	-106.6(18)	69.4(55)	-7.7(60)	57.9(38)
$\chi_1^{2,2}$	128.5(22)	-167.0(26)	178.5(23)	-175.9(18)
$\chi_2^1$	57.6(17)	57.0(31)	55.8(29)	55.7(32)
$\chi_{2}^{2.1}$	118.3(20)	105.4(29)	106.9(27)	91.0(44)
$\chi_{2}^{2,2}$	-121.3(17)	-127.5(32)	-129.3(25)	-161.3(32)
$\chi_3^1$	45.1(22)	58.2(25)	65.8(22)	38.6(25)
$\chi_{3}^{2,1}$	70.6(35)	97.0(31)	- 55.4(41)	77.4(28)
$\chi_3^{2,2}$	-172.3(26)	-159.9(24)	-179.9(19)	-165.7(24)
Xa1		171.8(18)	-173.1(32)	70.4(26)
$\chi_4^{2,1}$	_	73.8(28)	0.9(88)	77.7(31)
X4 <sup>2,2</sup>	_	-168.6(20)	160.8(45)	-155.2(25)

 $^{a} N^{3} - C_{a}^{3} - C_{I}^{3} - O^{T}$ .

planarity.<sup>24,25</sup> The  $\theta$  torsion angle of the *p*BrBz group, giving the orientation of the aromatic ring relative to the amide plane, exhibits values in the range  $\pm 26-30^{\circ}$ .<sup>21,22</sup> In particular, the sign for the three molecules of the tetramer is the same (positive).

The molecules of the  $N_{\alpha}$ -blocked tripeptide carboxylic acid pack into the unit cell in rows parallel to the **b** direction with intermolecular H-bonds between the amide N<sup>1</sup>-H and the peptide  $C_1^2=O^2$  of a symmetry-related molecule. The water molecule plays the role of the acceptor of the H-bond from peptide N<sup>2</sup>-H, while the donor of the H-bond to the acid  $C_1^3=O^3$  of a symmetry-related molecule.<sup>38,39</sup> In the crystals of the  $N_{\alpha}$ -blocked tetrapeptide ester molecules A form rows along the *c* direction with intermolecular H-bonds between the amide N<sup>1</sup>-H and the peptide  $C_1^3=O^3$  of a symmetry-related molecule A. The same observation applies to molecules B and C. No H-bonds between molecules of different types are seen.

#### Conclusions

In this work we have described the crystal-state structural tendency of two homochiral homopeptides (the trimer and tetramer) of ( $\alpha$ Me)Leu, as determined by X-ray diffraction. All ( $\alpha$ Me)Leu residues examined are found in the helical region of the conformational map. A 1 $\leftarrow$ 4C=O···H-N intramolecularly H-bonded  $\beta$ -turn conformation is adopted by the N<sup> $\alpha$ </sup>-blocked tripeptide carboxylic acid. This folded conformation is additionally stabilized by an unusual 1 $\leftarrow$ 4 C=O···H-O intramolecular H-bond, giving rise to an oxy-analogue of a  $\beta$ -turn at the C-terminus. The tetrapeptide ester forms a regular (incipient) 3<sub>10</sub>-helix with two 1 $\leftarrow$ 4 C=O···H-N intramolecular H-bonds. In addition, all D-( $\alpha$ Me)Leu residues (except those at the C-termini of the three independent molecules in the asymmetric unit of the tetramer) assume a right-handed helical conformation.

A comparison of the solid-state results described here for  $[(\alpha Me)Leu]_n$  homopeptides with the corresponding findings already reported for  $(Leu)_n$  homo-oligopeptides<sup>40-44</sup> allows us to conclude that the  $(\alpha Me)Leu$  residue is an effective  $\beta$ -turn and helix promoter, much stronger than its unmethylated parent compound Leu. Furthermore, in contrast to the Leu oligomers, there is no tendency for the  $(\alpha Me)Leu$  oligomers to adopt the self-associated  $\beta$ -sheet conformation.

The crystal-state preferred conformation of the ( $\alpha$ Me)Leu homo-oligomers, taken together with the corresponding tendency of the homopeptides derived from Aib (the prototype of  $C_n$ -methylated  $\alpha$ -amino acids),<sup>12-15</sup> Iva, ( $\alpha$ Me)Val, and

Table 2	Intra- and inter-molecular	H-bond parameters for	or pBrBz-[D-( $\alpha$ Me)Leu]	] <sub>3</sub> -OH monohydrate and	l pBrBz-[D-(aMe)Leu] <sub>4</sub> -OBu
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Peptide	Donor D-H	Acceptor A	Symmetry operations of A	Distance/Å D····A	Angle/degrees D··· A=C
(a) $pBrBz-[D-(\alpha Me)Leu]_3$ -OH monohydrate	N <sup>3</sup> -H	<b>O</b> <sup>0</sup>	<i>x</i> , <i>y</i> , <i>z</i>	2.93(1)	135.3(8)
	O'-H	$O^1$	<i>x</i> , <i>y</i> , <i>z</i>	2.65(1)	137.2(9)
	$N^1-H$	$O^2$	1 - x, -1/2 + y, 3/2 - z	3.03(1)	164.5(9)
	N <sup>2</sup> –H	Ow	x, 1 + y, z	3.08(2)	_
	O <sup>w</sup> –H	$O^3$	1 - x, -3/2 + y, 3/2 - z	2.88(2)	115(1)
(b) $pBrBz-[D-(\alpha Me)Leu]_4-OBu^t$	N <sup>3A</sup> –H	O <sup>0A</sup>	<i>x</i> , <i>y</i> , <i>z</i>	3.08(2)	131(1)
	N⁴^–H	O <sup>1A</sup>	x, y, z	3.33(2) <sup>a</sup>	132(1)
	N <sup>3B</sup> –H	O <sup>0B</sup>	x, y, z	2.85(2)	132(1)
	N <sup>4B</sup> –H	$O^{1B}$	x, y, z	$3.41(2)^{a}$	134(1)
	N <sup>3C</sup> –H	O <sub>0C</sub>	x, y, z	3.18(2)	135(1)
	N <sup>4C</sup> –H	O <sup>1C</sup>	x, y, z	3.18(2)	135(1)
	N <sup>1A</sup> –H	O <sup>3A</sup>	1 - x, 1 + (x - y), -1/3 + z	3.21(2)	155(1)
	N <sup>1B</sup> –H	O <sup>3B</sup>	1 + (y - x), 1 - x, 1/3 + z	3.16(2)	156(1)
	N <sup>1C</sup> –H	O <sup>3C</sup>	2 - x, 1 + (x - y), -1/3 + z	3.04(2)	153(1)

<sup>a</sup> Very weak interaction.

( $\alpha$ Me)Phe residues<sup>16</sup> reinforces the conclusions that  $C_{\alpha}$ methylation induces a marked propensity for  $\beta$ -turn and  $3_{10}$ helix formation in the resulting homo-oligomers.

As for the relationship between chirality of  $C_{\alpha}$ -methylated  $\alpha$ -amino acids and turn and helix handedness, the available experimental data point to the normal behaviour of protein amino acids (an L-residue prefers  $\varphi$ ,  $\psi$  backbone torsion angles typical of a right-handed helix) for the aliphatic Iva (with a linear side chain) and ( $\alpha$ Me)Val (with a  $\beta$ -branched side chain) residues,<sup>16</sup> but to a strong preference for an inverse relationship for the  $\gamma$ -branched aliphatic ( $\alpha$ Me)Leu (this work) and aromatic ( $\alpha$ Me)Phe<sup>16</sup> residues. In other words, the inverse relationship appears to be dictated by the position of side-chain branching (at the  $\gamma$ -carbon), rather than by the electronic nature (whether aliphatic or aromatic) of the side chain.

Not unexpectedly, the energy difference between right- and left-handed helical structures for  $C_{\alpha}$ -methylated chiral amino acids is much smaller than that of their protein counterparts.45-47 A widely accepted view is that the left-handed helical handedness of protein amino acids (L-configuration) is energetically disfavoured with respect to the diastereoisomeric right-handed helix mainly because of the close contact between side-chain  $C_{\beta L}$  atom and carbonyl oxygen of the same residue in the former conformation. Amino acids methylated at the  $\alpha\text{-carbon}$  possess two side-chain  $C_{\beta}$  atoms (a substituted  $C_{\beta L}$ atom and an unsubstituted  $C_{\beta D}$  atom). Since the helix handedness usually adopted by L-Iva (carrying a -CH2R substituent in the L side chain) is the same as that of  $L-(\alpha Me)Val$ [-CH(R)R] but opposite to that of  $L-(\alpha Me)Leu$  and L- $(\alpha Me)$ Phe [both of the -CH<sub>2</sub>-CH(R)R type], it is reasonable to assume that not only the preferred rotamers about the  $C_{\alpha}$ - $C_{\beta}$ bond ( $\chi^1$  torsion angles) but even those about the C<sub>B</sub>-C<sub>y</sub> bond, more removed from the backbone ( $\chi^2$  torsion angles), may play a role in directing  $\varphi$ ,  $\psi$  torsional preference of  $C_{\alpha}$ -methylated amino acids. To clarify this point, a statistical analysis relating backbone and side-chain torsion angles of the known X-ray diffraction structures of peptides containing  $C_{\alpha}$ -methylated amino acids<sup>16</sup> is currently in progress in our laboratory.

## References

- 1 G. Marshall, in Intra-Science Chemistry Report, ed. N. Kharasch, Gordon and Breach, New York, 1971, p. 305.
- 2 M. Mutter and S. Vuilleumier, Angew. Chem., Int. Ed. Engl., 1989, 28, 535.
- 3 I. L. Karle and P. Balaram, Biochemistry, 1990, 29, 6747.
- 4 C. Toniolo, F. Formaggio, M. Crisma, G. Valle, W. H. J. Boesten, H. E. Schoemaker, J. Kamphuis, P. A. Temussi, E. L. Becker and G. Précigoux, *Tetrahedron*, 1993, 49, 3641.
- 5 A. F. Spatola, in *Chemistry and Biochemistry of Amino Acids*, *Peptides and Proteins*, ed. B. Weinstein, Dekker, New York, 1983, vol. 7, p. 267.
- 6 V. J. Hruby, F. Al-Obeidi and W. Kazmierski, *Biochem. J.*, 1990, **268**, 249.
- 7 S. Polinelli, Q. B. Broxterman, H. E. Schoemaker, W. H. J. Boesten, M. Crisma, G. Valle, C. Toniolo and J. Kamphuis, *Bioorg. Med. Chem. Lett.*, 1992, 2, 453.
- 8 C. M. Venkatachalam, Biopolymers, 1968, 6, 1425.
- 9 C. Toniolo, CRC Crit. Rev. Biochem., 1980, 9, 1.
- 10 G. D. Rose, L. M. Gierasch and J. A. Smith, Adv. Protein Chem., 1985, 37, 1.
- 11 C. Toniolo and E. Benedetti, TIBS, 1991, 16, 350.
- 12 B. V. Venkataram Prasad and P. Balaram, CRC Crit. Rev. Biochem., 1984, 16, 1425.
- 13 C. Toniolo and E. Benedetti, *ISI Atlas of Science: Biochemistry*, 1988, 1, 225.

- 14 C. Toniolo, E. Benedetti and C. Pedone, Gazz. Chim. Ital., 1986, 116, 355.
- 15 C. Toniolo and E. Benedetti, Macromolecules, 1991, 24, 2004.
- 16 C. Toniolo, M. Crisma, F. Formaggio, G. Valle, G. Cavicchioni, G. Précigoux, A. Aubry and J. Kamphuis, *Biopolymers*, 1993, 33, 1061.
- 17 C. Toniolo, M. Pantano, F. Formaggio, M. Crisma, G. M. Bonora, A. Aubry, D. Bayeul, A. Dautant, W. H. J. Boesten, H. E. Schoemaker and J. Kamphuis, *Int. J. Biol. Macromol.*, 1994, in the press.
- 18 G. M. Sheldrick, SHELXS 86. Program for Crystal Structure Determination, University of Göttingen, Germany, 1986.
- 19 G. M. Sheldrick, SHELX 76. Program for Crystal Structure Solution and Refinement, Cambridge University, UK, 1976.
- 20 IUPAC-IUB Commission on Biochemical Nomenclature, J. Mol. Biol., 1970, 52, 1.
- 21 C. Toniolo, G. M. Bonora, A. Bavoso, E. Benedetti, B. Di Blasio, V. Pavone and C. Pedone, *Macromolecules*, 1986, **19**, 472.
- 22 A. Bavoso, E. Benedetti, B. Di Blasio, V. Pavone, C. Pedone, C. Toniolo, G. M. Bonora, F. Formaggio and M. Crisma, J. Biomol. Struct. Dynam., 1988, 5, 803.
- 23 W. B. Schweizer and J. D. Dunitz, Helv. Chim. Acta, 1982, 65, 1547. 24 E. Benedetti, in Chemistry and Biochemistry of Amino Acids, Peptides
- and Proteins, ed. B. Weinstein, Dekker, New York, 1982, vol. 6, p. 105. 25 T. Ashida, Y. Tsunogae, I. Tanaka and T. Yamane, Acta Crystallogr.,
- Sect. B, 1987, 43, 212. 26 S. S. Zimmerman, M. S. Pottle, G. Némethy and H. A. Scheraga,
- Macromolecules, 1977, 10, 1.
- 27 C. Toniolo and E. Benedetti, TIBS, 1991, 16, 350.
- 28 C. Toniolo, G. Valle, G. M. Bonora, M. Crisma, F. Formaggio, A. Bavoso, E. Benedetti, B. Di Blasio, V. Pavone and C. Pedone, *Biopolymers*, 1986, 25, 2237.
- 29 C. Ramakrishnan and N. Prasad, Int. J. Protein Res., 1971, 3, 209.
- 30 R. Taylor, O. Kennard and W. Versichel, Acta Crystallogr., Sect. B, 1984, 40, 280.
- 31 C. H. Görbitz, Acta Crystallogr., Sect. B, 1989, 45, 390.
- 32 J. Mitra and C. Ramakrishnan, Int. J. Peptide Protein Res., 1977, 9, 27.
- 33 I. D. Brown, Acta Crystallogr., Sect. A, 1976, 32, 24.
- 34 C. Toniolo, G. M. Bonora, A. Bavoso, E. Benedetti, B. Di Blasio, V. Pavone and C. Pedone, *Biopolymers*, 1983, **22**, 205.
- 35 E. Benedetti, G. Morelli, G. Némethy and H. A. Scheraga, Int. J. Peptide Protein Res., 1983, 22, 1.
- 36 R. O. Gould, A. M. Gray, P. Taylor and M. D. Walkinshaw, J. Am. Chem. Soc., 1985, 107, 5921.
- 37 J. Janin, S. Wodak, M. Levitt and B. Maigret, J. Mol. Biol., 1978, 125, 357.
- 38 C. H. Yang, J. N. Brown and K. D. Kopple, Int. J. Peptide Protein Res., 1979, 14, 12.
- 39 C. H. Görbitz and M. C. Etter, Int. J. Peptide Protein Res., 1992, 39, 93.
- 40 G. M. Bonora, A. Maglione and C. Toniolo, Polymer, 1974, 15, 767.
- 41 M. Palumbo, S. Da Rin, G. M. Bonora and C. Toniolo, *Makromol. Chem.*, 1976, **177**, 1477.
- 42 M. M. Kelly, E. S. Pysh, G. M. Bonora and C. Toniolo, J. Am. Chem. Soc., 1977, 99, 3264.
- 43 C. Toniolo, in *Bio-organic Chemistry*, ed. E. E. van Tamelen, Academic Press, New York, 1977, vol. 3, p. 265.
- 44 T. Ashida, I. Tanaka and T. Yamane, Int. J. Peptide Protein Res., 1981, 17, 322.
- 45 G. R. Marshall, J. D. Clark, J. B. Dunbar Jr., G. D. Smith, J. Zabrocki, A. S. Redlinski and M. T. Leplawy, *Int. J. Peptide Protein Res.*, 1988, 32, 544.
- 46 G. Valle, M. Crisma, C. Toniolo, R. Beisswenger, A. Rieker and G. Jung, J. Am. Chem. Soc., 1989, 111, 6828.
- 47 C. Toniolo, M. Crisma, G. M. Bonora, B. Klajc, F. Lelj, P. Grimaldi, A. Rosa, S. Polinelli, W. H. J. Boesten, E. M. Mejer, H. E. Schoemaker and J. Kamphuis, *Int. J. Peptide Protein Res.*, 1991, 38, 242.

Paper 3/05703H Received 21st September 1993 Accepted 2nd November 1993